



Research Article

www.ijrap.net



A COMPARATIVE ASSESSMENT OF PHYTOCHEMICAL SCREENING, ANTIOXIDANT AND ANTIMICROBIAL ACTIVITIES BETWEEN LEAF AND DUST OF BLACK TEA EXTRACTS

Shreya Mandal¹, Animesh Samanta¹, Arpita Patra¹, Shrabani Pradhan¹, Suchismita Roy¹, Koushik Das¹, Atiskumar Chattopadhyay², Dilip Kumar Nandi^{1*}

¹Department of Nutrition, Microbiology and Human Physiology, Raja N. L. Khan Women's College, Paschim Medinipore, West Bengal, India

²Secretary, Faculty Council of Science, Jadavpur University, Kolkata, West Bengal, India\

Received on: 14/09/16 Revised on: 07/10/16 Accepted on: 07/11/16

*Corresponding author

E-mail: dilipnandi2004@yahoo.co.in

DOI: 10.7897/2277-4343.076247

ABSTRACT

The present study is designed to investigate the comparison level of antimicrobial, antioxidant properties and as well as the total phenolic content estimation of aqueous extract between dusts and leaf black tea. Antimicrobial assay as well as antioxidant activity of phyto-compounds in both types of black tea was detected by using agar well diffusion and chromatographic technique respectively. Total antioxidant activity and phenolic content were measured by using colorimetric method. In comparison level the antimicrobial activity and screening of phytochemical compounds like saponins, phenolic compounds & tannins, flavonoids, triterpenoids were significantly ($P < 0.05$) different in both aqueous extract of leaf black tea and dust black tea. Total phenolic contents in the leaf black tea were 130.51mg Gallic acid equivalent/g of dried aqueous extract. In conclusion the leaf black tea has potential to be used as a natural antioxidant which is attributed to the rich presence of secondary metabolites and exhibit medicinal as well as physiological activities.

Keywords: Black tea, Phytochemicals, Antimicrobial activity, Antioxidant, Phenolic content, Chromatographic study

INTRODUCTION

Plants are the important source of drugs; especially in traditional medicine. In the world, phytomedicines had been used in past to treat various degenerative diseases long before the introduction of modern medicine. Medicinal plants are important for the treatment and management of human as well as animal diseases due to the presence of phytochemicals. Phytochemicals are naturally occurring compounds which are of great significance in the protection from various chronic diseases¹ and phytotherapy also is a cost effective and less side effect after treatment option. The most important bioactive constituents of plants are steroids, terpenoids, alkaloids, carotenoids, tannins, flavonoids, phenols and glycosides which serve as a valuable starting material for drug development².

Natural products can provide unlimited opportunities for the new drug discoveries because of their unmatched chemical diversity. The plant is contained natural active constituents which can be derived from any part of plant like flowers, bark, leaves, seeds, roots, fruits, etc. The useful medicinal effects of plant materials typically result from the combinations of the secondary products present in the plant³. Tea stands as the second most non-alcoholic beverage around the world and it is also the less expensive one⁴. Black tea, one of the best known of preparations, is made from *Camellia sinensis*⁵. Tea is generally consumed in the forms of green, black, and oolong tea, all of which are originated from the leaves of the plant *Camellia sinensis*. However the processing that leaves undergo makes different teas. The leaves for black tea are prepared by being fully oxidized before being dried. Tea leaves contain 10-30% of polyphenols including catechins, flavonols, phenolic acids, glycosides and plant pigments⁶. Tea leaves are a good source of polyphenols, specially thearubigins, theaflavins and catechins, a

decisive group for their antioxidative activity⁷⁻⁸. Tea polyphenols are also known by their antibacterial activity. In general, the antibacterial activity decreases when extent of the tea fermentation process, implies stronger activity in green tea than in black tea⁷. There are many number of reports on clinical uses of *Camellia sinensis* in various degenerative disorders particularly in cancer, tumor and diabetes that have shown promising results⁹. So, the present study aims to evaluate that the phytoconstituents, antioxidant and antimicrobial activities are compared between the leaf and dust black teas of aqueous extracts.

MATERIALS AND METHODS

Plant materials and chemicals

The branded leaf and dust black tea were collected from open market at Midnapur district, West Bengal, India. The materials were identified by the taxonomist of the Botany Department at the Raja N. L. Khan Women's College, Midnapore. The voucher specimens (Leaf Tea-DJ/497/07042015/TE, Dust Tea-10013022001897) were deposited in the Department of Botany, Raja N. L. Khan Women's College at Paschim Medinipur, West Bengal. All other chemicals used were of analytical grade and were purchased from HiMedia Laboratories Pvt. Limited (Mumbai, India), SRL (India).

Bacterial strain and culture conditions

Two gram negative and two gram positive indicator bacteria are used for antimicrobial assay respectively *Escherichia coli* (MTCC443), *Klebsiella Pneumoniae* (MTCC 109), *Staphylococcus aureus* (MTCC 3160), and *Streptococcus mutans* (MTCC890), provided by Microbiological laboratory and clinical detection center Midnapur (Paschim Medinipur,

India). They were cultured in tryptone soy broth or agar (TSB or TSA) in aerobic condition at 37 °C.

Black tea extracts preparation

Black tea extracts were prepared by adding leaf tea (LT) and dust tea (DT) in 25gm to 250mL of boiling water in each conical flask, steeped for 15–20 min. The infusion was cooled at room temperature and then filtered through Whatman No.1 filter paper. Both resulting filtrate were dried in the air, weight and stored in air tight vacuum container for different analysis.

Phytochemicals analysis

Phytochemicals analyses of the test samples were carried out according to standard methods¹⁰⁻¹².

Test for phytosterols- Salkowski reaction

About 0.5 ml chloroform was added to both extracts in test tubes. Then 1ml of Conc. H₂SO₄ was added to it from the sides of the test tube. Appearance of reddish brown colour in chloroform layer indicates presence of phytosterols.

Test for triterpenoids- Liebermann - Burchard's test

Extracts were treated with few drops of acetic anhydride, boiled and cooled and then Conc. Sulfuric acid was added from the sides of the test tube. A brown ring was shown at the junction of two layers and formation of deep red colour and indicates the presence of triterpenoids.

Test for saponins- Foam test

Small amount of both extracts were taken in test tubes with little quantity of water. Shake vigorously. Appearance of foam which is persisting for 10 minutes indicates presence of saponins.

Test for alkaloids- Dragendorff test

Dissolve both the extracts of black teas by chloroform. Evaporate chloroform and acidify the residue by adding few drops of Dragendorff reagent (Potassium Bismuth Iodide). Appearance of orange red precipitation indicates the presence of alkaloids.

Test for carbohydrates- Molisch's test

Mix the extracts with Molisch's reagent and add Conc. H₂SO₄ along the sides of the test tube. Appearance of reddish violet ring the interference indicates the presence of carbohydrates.

Test for flavonoids- Lead acetate test

The alcoholic solution of the extracts was added to few drops of 10% Lead acetate solution. Appearance of yellow precipitation indicates the presence of flavonoids.

Test for phenolic compounds and tannins- Ferric chloride test

Both extracts (each 2ml) was taken in the test tubes and ferric chloride solution was added to it drop by drop. Appearance of bluish black precipitation indicates the presence of phenolic compounds and tannins.

Test for proteins- Ninhydrin test

Few drops of Ninhydrin are added to the both extracts. Appearance of blue colour indicates presence of amino acid whereas proteins may rarely give positive result.

Test for glycosides- Keller-Killiani test

Both extracts were taken in the test tubes, to add 1ml of glacial acetic acid and few drops of ferric chloride solution and also Conc. H₂SO₄ in each extract (Slowly through the sides of the

test tube). Appearance of reddish brown ring at the junction of the two liquids indicates the presence of de-oxy-sugars.

Thin layer chromatography analysis for antioxidant constituents

About 2 µg of both extracts of tea were loaded on a TLC plate (Merck, 20 cm x 20 cm). The plate was developed with methanol: chloroform: hexane (7:2:1, v/v/v) to separate various constituents of the extracts. The developed plates were dried by hair drier. Then the antioxidant constituents were analyzed by DPPH technique. For this purpose, 0.05% of methanolic solution of DPPH was sprayed on the surface of developed TLC plates and incubated for 10 min at room temperature. The active antioxidant constituents of both the extracts of tea were detected as yellow colour was produced by the reduction of DPPH in the purple background on the TLC plates. Ascorbic acid was used as standard antioxidant compound¹³.

Determination of total phenol content

The amounts of phenol compounds in both the extracts of tea were determined using folin ciocalteu reagent, according to the modified method¹⁴. 1 ml of the plant extract/standard solution was mixed with 5 ml Folin-Ciocalteu reagent and 4 ml (7.5% sodium carbonate) of sodium carbonate (Na₂CO₃). The tubes were vortexed for few seconds and allowed to stand for 30 min at 20°C for colour development. Absorbance of samples and standard were measured at 765 nm using spectrophotometer against blank. A typical blank solution contained the solvent used to dissolve the plant extract. The total content of phenolic compounds plant extracts in gallic acid equivalents (GAE) was calculated using the following equation:

$$C = (c \times V)/m$$

Where C = total content of phenolic compounds, mg/g plant extract, in GAE, c = the concentration of gallic acid established from the calibration curve (mg/ml), V = the volume of extract in ml, and m = the weight of plant extract in g.

Antioxidant activity determination by DPPH free radical scavenging assay

DPPH free radical scavenging activity of both the extracts of tea was measured by this method¹⁵. For this analysis, ascorbic acid (Standard) was dissolved in methanol (Sigma-Aldrich) and methanol fractions of TA bark were used as the test solutions. About 1 ml of each fraction was placed into test tubes and 0.5 ml of 1 mmol/L DPPH solution in methanol was added. The test tubes were incubated for 15 min and the absorbance was read at 517 nm. A blank solution contained of DPPH dissolved in same amount of methanol. The DPPH radical scavenging activity percentage was calculated by using the following formula:

$$\text{DPPH radical scavenging activity (\%)} = \frac{[A_{517\text{control}} - A_{517\text{extract}}] / A_{517\text{control}} \times 100}$$

Antimicrobial analysis

The antimicrobial activity was determined in both extracts of tea using agar well diffusion method. The antibacterial activities of both black tea aqueous extracts (concentration of compound 50%, 100 %) were tested against two Gram-positive— *S. aureus*, *S. mutans* and two Gram-negative— *E. coli* and *K. pneumoniae*, human pathogenic bacteria; Zone of inhibition of both black tea aqueous extracts were compared with standards like chloramphenicol for antibacterial activity. The results

showed that the remarkable inhibition of the bacterial growth was shown against the tested organisms¹⁶.

Statistical Analysis

The values were expressed as Mean ± SE. Data were analyzed using one-way ANOVA followed by t-test. P value < 0.05 was considered as significant¹⁷. ANOVA was followed by multiple two-tail t-test and data with different superscripts (a, b) in a specific vertical column differ from each other significantly (P< 0.05). Statistical analysis was performed using SPSS 12.0 and MS-Excel 2007.

RESULTS

The result obtained from gram percent (g %) of aqueous extract of both the leaf and dust black teas were 66g% and 43.6g% respectively as shown in table 1. So, the g% of aqueous extract of leaf of black tea was greater than dust of black tea. The preliminary phytochemicals screening was revealed that the presence of saponins, phytosterols, triterpenoids, alkaloids, carbohydrates, polyphenols, tannins, proteins, glycosides and flavonoids were compared between leaf and dust of black tea of aqueous extracts as shown in the table 2. In this experiment some secondary metabolites (such as saponins, triterpenoids, polyphenols, flavonoids and tannins) were present in high concentration of aqueous extract of black leaf tea than black dust tea.

The plates TLC were developed in methanol: chloroform: hexane (7:2:1, v/v/v) and sprayed with 0.05% DPPH reagent. Purple colour of DPPH reagent was bleached by yellow spots was the indication of positive antioxidant activity. Both the aqueous extracts of dust and leaf of black teas in terms of DPPH free radical scavenging activity showed one resolved TLC band with strong antioxidant activity and another spot with weak antioxidant activity as compared to standard antioxidant ascorbic acid. In this experiment both types of black tea showed antioxidant activities in comparison to standard ascorbic acid (Figure 1). DPPH is a free radical and it gives strong absorption band at 517 nm in the visible region of electromagnetic radiation. As antioxidant compounds donate protons to these radicals, the absorption decreases. The decrease in absorption is taken as a measure of the extent of radical scavenging activity. The results were compared with that of ascorbic acid and the aqueous extracts of both black teas. The leaf of black tea showed better result than dust of black tea (Figure 2). Total phenolic content of both teas of aqueous extracts was shown in table 4 and figure 3. In this experiment black of leaf tea was contained huge amount polyphenols than dust of black tea. They were significantly (P< 0.05) differ from each other. Antimicrobial activity of both black teas aqueous extracts showed greater result against gram negative bacteria than gram positive bacteria as shown in table 3. In this experiment the aqueous extract of leaf and dust black tea showed that greater inhibition zone against gram negative bacteria than gram positive bacteria. But leaf black tea showed better antimicrobial activity than dust black tea (Figure 4).

Table 1: Percentage of extract from aqueous of the leaf and dust black teas

| Black Tea | Amount of Solvent (ml) | Amount of Tea (g) | Amount of extract (g) | Percentage of extract (g %) |
|-----------|------------------------|-------------------|-----------------------|-----------------------------|
| BLT | 250 | 25 | 16.5 | 66 |
| BDT | 250 | 25 | 10.9 | 43.6 |

BLT: Black Leaf Tea, BDT: Black Dust Tea

Table 2: Preliminary phytochemicals analysis of the leaf and dust black teas of aqueous extracts

| Sl. No. | Phytoconstituents | Tests | BLT | BDT |
|---------|------------------------------|------------------------------|-----|-----|
| 1. | Phytosterols | Salkowski reaction | + | + |
| 2. | Triterpenoids | Liebermann - Burchard's test | ++ | + |
| 3. | Saponins | Foam test | ++ | + |
| 4. | Alkaloids | Dragendroff's test | + | - |
| 5. | Carbohydrates | Molisch's test | - | - |
| 6. | Flavanoids | Lead Acetate test | ++ | + |
| 7. | Phenolic Compounds & Tannins | 5% FeCl ₃ Test | ++ | + |
| 8. | Proteins | Ninhydrin test | - | - |
| 9. | Glycosides | Keller-Killiani test | - | - |

BLT: Black Leaf Tea, BDT: Black Dust Tea, ++ : Highly present, + : Slightly present, - : Absent

Table 3: Antimicrobial activities of the leaf and dust black tea aqueous extracts and zone of inhibition

| | Diameter of Zone of inhibition (mm)* against | | | | | | | |
|-----------------------|--|-----|-----------------------------|-----|------------------------------|-----|-------------------------|-----|
| | <i>Staphylococcus aureus</i> | | <i>Streptococcus mutans</i> | | <i>Klebsiella pneumoniae</i> | | <i>Escherichia coli</i> | |
| | Conc. of compound | | Conc. of compound | | Conc. of compound | | Conc. of compound | |
| | 100% | 50% | 100% | 50% | 100% | 50% | 100% | 50% |
| BLT | 15 | 7.8 | 17 | 8.2 | 23 | 8.5 | 35 | 22 |
| BDT | 9.7 | 4.6 | 13 | 5.8 | 17 | 5.5 | 23 | 10 |
| Chloramphenicol (STD) | 18 | 10 | 20 | 12 | 28 | 15 | 45 | 25 |

*The zone of inhibition (mm) taken as average, BLT: Black Leaf Tea, BDT: Black Dust Tea, STD: Standard

Table 4: Total phenolic content estimation from the leaf and dust of black tea aqueous extracts (mg GAE/g). Data are expressed as Mean ± SE (n=6)

| Black Tea Aqueous Extract | Total phenol contents (mg/g, Gallic Acid Equivalent) |
|---------------------------|--|
| BLT | 130.51±0.41 ^a |
| BDT | 82.42±0.85 ^b |

BLT: Black Leaf Tea, BDT: Black Dust Tea



Figure 1: Thin layer chromatography antioxidant activity analysis of Black tea water extracts and Ascorbic acid. TD- Tea Dust, TL- Tea Leaf, STD- Ascorbic acid

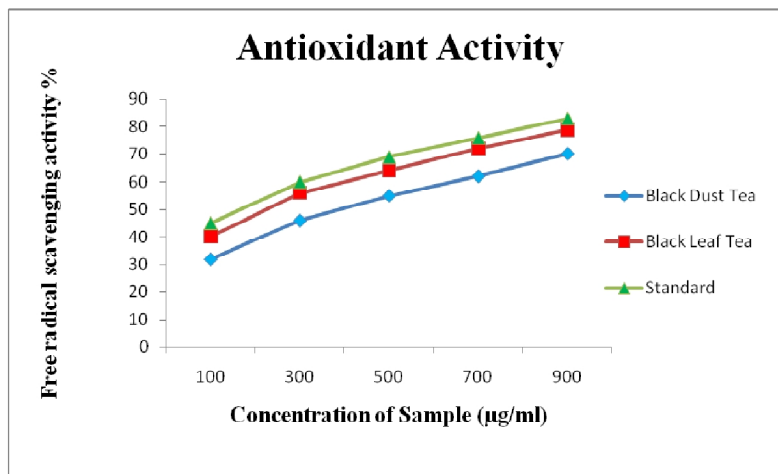


Figure 2: DPPH free radical scavenging activity of Black tea dust and leaf water extracts Standard- Ascorbic acid

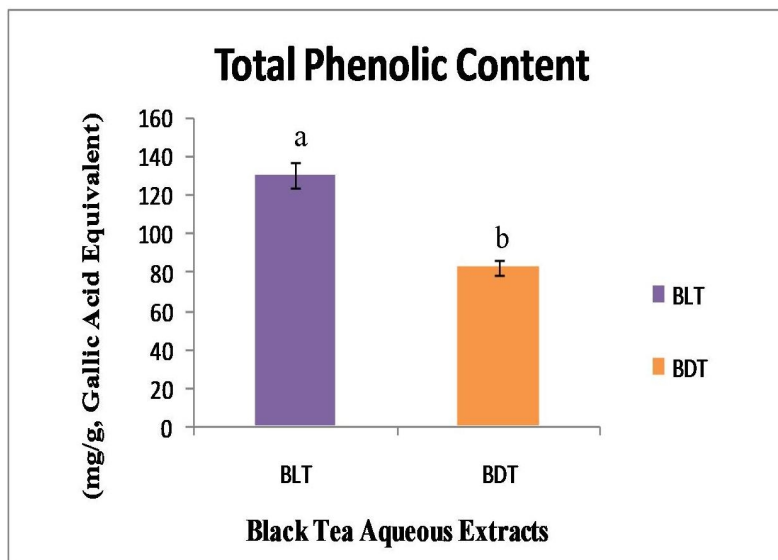
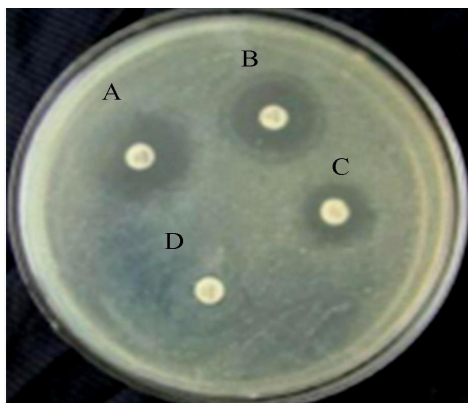


Figure 3: Total phenol contents of the aqueous extracts of BLT and BDT. Data are expressed as mean \pm SE (n=6). BLT: Black Leaf Tea, BDT: Black Dust Tea



**Figure 4: Inhibition zone against *Escherichia coli* (Indicator microbes of gram negative bacteria)
A-BLT (Black Leaf Tea), B- Chloramphenicol (STD), C- BDT (Black Dust Tea), D- Control (Aqueous)**

DISCUSSION

In the present investigation, the obtained gram percent (g%) of aqueous extract of both the leaf and dust black teas revealed that black leaf tea aqueous extract contained huge amount of phytochemicals and the extracted quantity of black leaf tea was greater than dust black tea. Physicochemical standardization is the most prominent means for quality assurance of herbal products¹⁸. Phytochemical analysis conducted on the tea extracts revealed the presence of constituents which are known to exhibit herbal medicinal as well as physiological activities¹⁹. The aqueous extract of leaf of black tea showed high concentration of saponins, triterpenoids, polyphenols, flavanoids and tannins than aqueous extract of dust of black tea. Due to presence of huge amount of phytoconstituents in leaf of black tea, it showed higher antioxidant activity than dust of black tea. This antioxidant activity result in both teas extracts was shown in figure 1 and 2. This is related to the fact that antioxidants can prevent free radicals, primarily highly reactive oxygen and nitrogen species, from damaging human health. Harmful effects of free radicals and oxidative stress can be reduced by regular consumption of tea like beverages which is exhibits antioxidant activity²⁰. Phytochemicals are known to be synthesized by plants in response to different microbial infection and they have been found to be different antimicrobial substances against the wide array of micro-organisms in-vitro and their activities are probably due to their ability to complex with extracellular and soluble proteins and to complex with the bacterial cell wall²¹. The antimicrobial activity of leaf and dust of black tea was detected against four entero-pathogenic bacterial strains: *Staphylococcus aureus*, *Streptococcus mutans*, *Klebsiella pneumonia*, *Escherichia coli*, and compared to that of reference Standard drug Chloramphenicol disc. Aqueous extract of both teas showed inhibition against tested strains to varying degrees of 100% and 50% concentrations. The black tea leaf (BLT) of aqueous extract showed maximum inhibition against tested strain. Thus, the aqueous extract of BLT has shown great antioxidant and antimicrobial activities. Phenolic compound embrace a broad range of plants secondary metabolite which have health favourable properties. The accumulation of laboratory and clinical studies suggested that polyphenol-rich plants have health promoting effects with respect to metabolic health²² and cancer prevention²³. Polyphenols are natural antioxidant from plants and are consumed in the forms of vegetables, fruits and beverage such as tea, coffee and wine²⁴. The known in-vitro antioxidant properties of catechins and other polyphenolic compounds of tea have led to interest in the potential health benefits of tea consumption^{25,26}. In this

experiment, there is a significant ($P < 0.05$) difference between in leaf and dust of black tea aqueous extracts. Black leaf tea contained more amount of phenolic content than black dust tea. This result denoted that the leaf of black tea possess polyphenols, chiefly responsible for antioxidant activity. It was mentioned that the presence of phenolic content could show the antioxidant and free radical scavenging properties which was confirmed by our experiment. According to our investigation, the high contents of polyphenol in the leaf of black tea can explain its high free radical scavenging activity. The results obtained from the above experiment showed that leaf of black tea contains different major antioxidative compounds in high concentration which may be helpful for treatment of diseases as well as for suppressing the growth of many pathogenic organisms.

CONCLUSION

It has been shown that leaf of black tea consists of many useful compounds such as flavonoids, tannins, phytosterols, saponins, polyphenols and alkaloids. Its antioxidant activity is largely due to the presence of polyphenols. Tea polyphenols or total phenolic contents are well-known for their antioxidant properties. The antioxidant and antimicrobial properties of black leaf tea are responsible on presence of large amount of phytochemicals. So the results further support the view that the black leaf tea is a promising source of natural useful therapeutic agents. Further, there is a need to isolate, purify and identify these natural active constituents present in leaf black tea as an important medicinal plant, which is extensively used in Indian system of medicine for prevention of degenerative diseases.

ACKNOWLEDGMENT

We are thankful to the honourable Principal Dr. Jayasree Laha Raja N. L. Khan Women's College, Paschim Medinipur of her infrastructural support and Dr. Joyjit Ghosh, Dept. of English, Vidyasagar University for his kind cooperation in correction of English language.

REFERENCES

1. Geoffrey KK, John KM, Naomi M, Simon KM. Qualitative phytochemical screening of *Camellia sinensis* and *Psidium guajava* leave extracts from Kericho and Baringo counties. International Journal of Advanced Biotechnology and Research 2014; 15(3): 506-512.

2. Tariq AL, Reyaz AL. Phytochemical analysis of *Camellia sinensis* leaves. International Journal of Drug Development and Research 2012; 4(4):311-316.
3. Makari H, Haraprasad N, Ravikumar P. Vitro antioxidant activity of the hexane and methanolic extracts of *Cordia wallichii* and *Celastrus paniculata*. The Internet Journal of Aesthetic and Antiaging Medicine 2007; 1(1): 1-10.
4. Sharangi AB. Medicinal and therapeutic potentialities of tea [*Camellia sinensis* L.]. Food Research International 2009; 42: 529-535.
5. Funmilayo OO, Kamaldeen AS, Buhari ASM. Phytochemical screening and antimicrobial properties of a common brand of black tea (*Camellia sinensis*) marketed in Nigerian environment. Advanced Pharmaceutical Bulletin 2012; 2(2): 259-263.
6. Pan X, Niu G, Liu H. Microwave-assisted extraction of tea polyphenols and tea caffeine from green tea leaves. Chemical Engineering and Processing 2003; 42(2): 129-133.
7. Kaur HP, Kaur S, Rana S. Antibacterial activity and phytochemical profile of green tea, black tea and divya peya herbal tea. International Journal of Pure Applied Bioscience 2015; 3(3): 117-123.
8. Salah N, Miller NJ, Parganga G, Tifburg L, Bolwell GP, Ice-Evan C. Polyphenolic flavonols as scavengers of aqueous phase radicals and as chain-breaking antioxidants. Arch Biochem Biophys 1995; 322(2): 339-346.
9. Gupta V, Bansal P, Niasi J, Kumar S. Phytochemistry and pharmacology of *Camellia sinensis*. Annals of Biological Research 2010; 1(2): 91-102.
10. Harborne Jb. Phytochemical methods: A guide to modern techniques of plant analysis. 3rd ed. London: Chapman and Hall; 1998.
11. Fransworth NR. Biological and phytochemical screening of plants. Journal of Pharmaceutical Science 1966; 55(3): 225-267.
12. Rangari VD. Pharmacognosy and Phytochemistry. 1st ed. Nasik: Carrier Publication; 2002.
13. Mandal S, Patra A, Samanta A, Roy S, Mandal A, Pradhan S et al. Analysis of phytochemical profile of *Terminalia arjuna* bark extract with antioxidative and antimicrobial properties. Asian Pac J Trop Biomed 2013; 3(12): 960-966.
14. Demiray S, Pintado ME, Castro PML. Evaluation of phenolic profiles and antioxidant activities of turkish medicinal plants: *Tilia Argentea*, *Crataegi folium* leaves and *Polygonum bistorta* roots. World Academy of Science, Engineering and Technology 2009; 54:312-317.
15. Barros L, Falcao S, Baptista P, Freire C, Vilas-Boas M, Ferreira IC. Antioxidant activity of *Agaricus sp.* mushrooms by chemical, biochemical and electrochemical assays. Food Chemistry 2008; 111:61-66.
16. Nema R, jain P, Khare S, Pradhan A, Gupta A, Singh D. Antibacterial and antifungal activity of *Terminalia Arjuna* leaves extract with special reference to flavonoids. Basic Research Journal of Medicine and Clinical Sciences 2012; 1(5): 63-65.
17. Pradhan S, Mandal S, Roy S, Mandal A, Das K, Nandi DK. Attenuation of uremia by orally feeding alpha-lipoic acid on acetaminophen induced uremic rats. Saudi Pharma J 2013; 21(2):187-192.
18. Sharma PK, Ali M, Yadav DK. Physicochemical and Phytochemical evaluation of different black tea brands. Journal of Applied Pharmaceutical Science 2011; 01 (03): 121-124.
19. Sofowra A. Medicinal Plants And traditional Medicine in Africa. 2nd ed. Nigeria: Spectrum Books Ltd; 1993.
20. Yashin A, Yashin Y, Nemzer B. Determination of antioxidant activity in tea extracts, and their total antioxidant content. Am J Biomed Sci 2011; 3(4): 322-335.
21. Cowan MM. Plant products as antimicrobial agents. Clinical Microbiol Rev 1999; 12(4): 564-582.
22. Chen N, Bezzina R, Hinch E, Lewandowski PA, Cameron-Smith D, Mathai ML et al. Green tea, black tea and epigallocatechin modify body composition, improve glucose tolerance and differentially alter metabolic gene expression in rats fed a high-fat diet. Nutr Res 2009; 29(11): 784-793.
23. Ferguson PJ, Kurowska E, Freeman DJ, Chambers AF, Koropatnick DJ. A flavonoid fraction from cranberry extract inhibits proliferation of human tumor cell lines. J Nutr 2004; 134(6): 1529-1535.
24. Uchenna JU, Selena A, Adam K, James TL, Edward JK. White and green teas (*Camellia sinensis var. sinensis*): Variation in phenolic, methylxanthine and antioxidant profiles. J Food Sci 2010; 75(6): 66-78.
25. Higdon JJ, Frei B. Tea catechins and polyphenols: health effects, metabolism, and antioxidant functions. Crit Rev Food Sci Nutr 2003; 43(1): 89-143.
26. Abhishek Kumar, Anil Kumar Singh, Surendra Singh. Extraction, phytochemical screening, separation and characterisation of bioactive compounds from leaves extracts of *Clitoria ternatea* Linn. (Aparajita). Int. J. Res. Ayurveda Pharm. Sep - Oct 2016;7(5):70-77 <http://dx.doi.org/10.7897/2277-4343.075198>

Cite this article as:

Shreya Mandal, Animesh Samanta, Arpita Patra, Shrabani Pradhan, Suchismita Roy, Koushik Das, Atiskumar Chattopadhyay, Dilip Kumar Nandi. A comparative assessment of phytochemical screening, antioxidant and antimicrobial activities between leaf and dust of black tea extracts. Int. J. Res. Ayurveda Pharm. Nov - Dec 2016;7(6):90-95 <http://dx.doi.org/10.7897/2277-4343.076247>

Source of support: Nil, Conflict of interest: None Declared

Disclaimer: IJRAP is solely owned by Moksha Publishing House - A non-profit publishing house, dedicated to publish quality research, while every effort has been taken to verify the accuracy of the content published in our Journal. IJRAP cannot accept any responsibility or liability for the site content and articles published. The views expressed in articles by our contributing authors are not necessarily those of IJRAP editor or editorial board members.